3 to 1

4. Cagampang Ramos, Adela and Richard F. Darsie, Jr. 1970 (May). Illustrated Keys to the Anopheles mosquitoes of the Philippine Islands. USAF Fifth Epidemiological Flight, PACAF, Tech. Rept. 70-1. 49 pp. Address: APO San Francisco, California 96528.

This is a beautifully illustrated publication dealing with the identification of the adult females and fourth instar larvae of Philippine Anopheles. It is dedicated to Francisco Edlagan Baisas, fittingly described as "Dean of Philippine Culicidologists." Forty species and subspecies of Anopheles are included.

5. Ault, P. 1970. Wonders of the mosquito world. Dodd, Mead and Company, N. Y. 64 pp. \$3.50.

This interesting little volume is one of a series entitled "Wonders of the ____ " and is specifically written for the general public. It is worth noting here for two reasons, however. First, it will be of use wherever there is a need for training individuals to participate in mosquito control programs and second, it is dedicated to Dr. George B. Craig, Jr. who has contributed so much to mosquito systematics via the avenue of mosquito genetics.

Illustrated KEYS to the **ANOPHELES** MOSQUITOES of the **Philippine** Islands

USAF FIFTH EPIDEMIOLOGICAL FLIGHT, PACAF **TECHNICAL REPORT 70-1** APO SAN FRANCISCO 96528 **MAY 1970**

BY

RICHARD F. DARSIE, JR. MALARIA ERADICATION TRAINING CENTER MANILA, PHILIPPINES

Fage 3, column 1, line 27 - for Delfinado (1965) ry Page 6, column 1, line 16 - for halteres, read he

Page 30, column 1, line 21 - for Fig. 102 reag

Page 30, column 1, line 41 - for however, Vrage 30, column 2, line 43 - for serration 176.

Page 46, Figure 177 - Place letter A vunder righthand drawing.

Page 46, Footnote, line 2 - delete Page 48, line 24 - for Monthly Bull

USAF Fifth Epidemiological Flight, PACAF

Technical Report 70-1

ILLUSTRATED KEYS TO THE ANOPHELES MOSQUITOES OF THE PHILIPPINE ISLANDS

by

ADELA CAGAMPANG-RAMOS, B. Sc. Agr. Entomologist, Malaria Eradication Training Center

and

RICHARD F. DARSIE, JR., Ph. D. Malaria Training Advisor (Entomology) U. S. Public Health Service

USAF Fifth Epidemiological Flight, PACAF

APO San Francisco 96528 and

Malaria Eradication Training Center Manila, Republic of the Philippines

May 1970

The opinions expressed herein are those of the authors and do not necessarily represent those of the United States Air Force.

To

FRANCISCO EDLAGAN BAISAS Dean of Philippine Culicidologists

we respectfully dedicate this work. His contribution to the knowledge of Philippine mosquitoes

Table of Contents

1	Table of Key Figures
	Introduction
	Classification of Philippine Anopheles
1	Systematic Index
	Morphology of Adult Female
	Head
	Thorax
	Abdomen
	Illustrated Key to Adult Females of Philippine Anopheles
	Morphology of Fourth Instar Larva
	Head
	Thorax
	Abdomen
	Illustrated Key to Fourth Instar Larvae of Philippine Anopheles 31
	References

Acknowledgments

The authors are very much indebted to Mr. F.E. Baisas, who contributed substantially to the formation of the keys and loaned several illustrations, and to Dr. H.F. Kroening, Senior Malaria Training Advisor, U.S. Public Health Service, Malaria Eradication Training Center, Manila, Philippines, for his support of the study.

They are deeply grateful to the Commanding Officers of the Fifth Epidemiological Flight (PACAF), U.S. Air Force, and the 406th Medical Laboratory, Department of the Army, U.S. Department of Defense, and to the staff entomologists of these Units: Major Dale W. Parrish, Capt. Stephen Valder, Lt. Col. Vernon Tipton and Lt. Col. A.A. Hubert for their support of the project. Most of the figures were prepared by the artists of the 406th Medical Laboratory, and the work of Kei Daishoji, Sei Fujisawa, Tadashi Tanami, Saburo Shibata, Masao Hasunuma, Ichiro Yoshigaki and their supervisor, Miss Yoshika Yoshida is hereby acknowledged. In addition, some of the figures were prepared by Mr. Angel Martinez and by Mr. Alfredo Danganan; they also assisted with the innumerable details in finalizing all the illustrations. Thanks go to Miss Dalisay M. del Rosario and Mr. Ramon L. Reyes, Jr. for their part in the preparation of the many drafts.

The loan of specimens from the U.S. National Museum, through the courtesy of Drs. Alan Stone and Botha DeMeillon, is appreciated.

Table of Key Figures

ADULTS

- 7. Wing of A. peditaeniatus
 8. Wing of A. m. flavirostris
- 9. Fore femur of A. franciscoi
- 0. Proboscis and palpi of A. manalangi
- 11. Fore femur of A. aitkenii
- 12. Fore femur of A. I. benguetensis13. Proboscis and palpi of A. I. benguetensis
- Proboscis and palpi of A. L benguetensi.
 Proboscis and palpi of A. manalangi
- 15. Proboscis and palpi of A. peditaeniatus
- 16. Abdominal sternites of A. samarensis
- 17. Abdominal sternites of A. franciscoi18. Hind leg of A. samarensis
- 19. Hind leg of A. balerensis (Redrawn
- from Mendoza, 1948) 20. Same as 19
- 21. Hind leg of A. ejercitoi (Redrawn from Mendoza, 1948)
 - 2. Fore femur of A. pseudobarbirostris
 - . Abdominal sternites of A. pseudobarbirostris
 . Same as 9
- 25. Same as 17
 - . Wing of A. franciscoi
- 27. Wing of A. vanus28. Thoracic pleuron of A. vanus
- 29. Thoracic pleuron of A. manalangi
- 30. Hind tarsomeres of A. peditaeniatus
 31. Hind tarsomeres of A. lesteri
- 31. Hind tarsomeres of A. lesteri
 32. Thoracic pleuron of A. pseudosinensis
- 33. Wing base of A. pseudosinensis
- Thoracic pleuron of A. lesteri
 Wing base of A. lesteri
- Wing of A. aitkenii
 Hind leg of A. aitkenii
- 38. Wing of A. g. formosus
- 39. Hind leg of A. I. benguetensis40. Hind femur of A. I. benguetensis
- 41. Wing of A. L benguetensis42. Hind femur of A. g. formosus
- 43. Same as 38
- 44. Hind tarsomeres of A. maculatus
 45. Hind tarsomeres of A. kochi
- 46. Hind leg of A. maculatus
- 47. Hind leg of A. karwari
 48. Hind tarsomeres of A. karwari

- 49. Proboscis and palpi of A. karwari
- 50. Hind tarsomeres of A. annularis51. Proboscis and palpi of A. annularis
- 52. Wing of A. annularis
- 53. Wing of A. philippinensis
 54. Wing of A. b. balabacensis
- 55. Abdominal tergites of A. kochi
- 56. Wing of A. parangensis57. Abdominal tergites of A. b. balabacensis
- 58. Same as 55 59. Same as 57
- 60. Hind leg of A. b. balabacensis
 61. Hind leg of A. tessellatus
- 62. Same as 54
 63. Hind tibio-tarsal joint of A. b. balabacensis
- 63. Hind fibio-tarsal joint of A. b. balabace (Redrawn from Colless, 1956) 64. Wing of A. cristatus (Redrawn from
- King and Baisas, 1936)
 65. Hind tibio-tarsal joint of A. r. riparis
- (Redrawn from Colless, 1956) 66. Same as 54
- 67. Same as 6068. Wing of A. b. baisasi
- 69. Hind leg of A. b. baisasi70. Same as 64
- 71. Wing of A. r. riparis (Redrawn from
- King and Baisas, 1936)
 72. Abdominal tergites of A. tessellatus
- 73. Proboscis and palpi of A. tessellatus74. Abdominal tergites of A. kolambuganensis
- 74. Abdominal tergites of A. kolambuganensis
 75. Proboscis and palpi of A. kolambuganensis
- 77. Wing of A. v. limosus
- 78. Wing of A. filipinae
 79. Proboscis and palpi of A. filipin
- 79. Proboscis and palpi of A. filipinae
- 81. Proboscis and palpi of A. m. flavirostris
- 82. Same as 81 83. Same as 8
- 84. Proboscis and palpi of A. mangyanus
 - 85. Wing of A. mangyanus86. Hind leg of A. I ludlowae
 - 86. Hind leg of A. I. hidlowae 87. Hind leg of A. v. limosus
 - 88. Hind tarsomeres of A. parangensis
 (Redrawn from Russell and Baisas, 1936)

- 89. Hind tarsomeres of A. I. ludlowae
- 90. Hind femur and tibia of A, litoralis
- 91. Hind femur and tibia of A. l. ludlowae
- 92. Wing of A. L. ludlowae
- 93. Wing of A. I. cabrerai 94. Proboscis and palpi of A. v. limosus
- 95. Same as 77
- 96. Proboscis and palpi of A. indefinitus
- 97. Wing of A. indefinitus
- 98. Proboscis and palpi of A. v. vagus
- 99. Wing of A. v. vagus
- 99a. Same as 94
- 99b. Same as 77
- 99c. Proboscis and palpi of A. subpictus
- 99d. Same as 96

LARVAE

- 105. A Clypeal hairs of A. franciscoi B Antenna of A. franciscoi
- 106. A Clypeal hairs of A. litoralis B Antenna of A. litoralis
- 107. Clypeal hairs of A. g. formosus 108. Clypeal hairs of A. pseudobarbirostris
- 109. Abdominal segment I of A. g. formosus
- 110. Abdominal segment I of A. insulaeflorum
- 111. Abdominal palmate on III of A. g. formosus
- 112. Abdominal palmate on III of A. l. benguetensis
- 113. Clypeal hairs of A. insulaeflorum
- 114. Abdominal segment III of A. insulaeflorum
- 115. Clypeal hairs of A. bengalensis
- 116. Abdominal segment III of A. bengalensis
- 117. Clypeal hairs of A. fragilis
- 118. Clypeal hairs of A. aitkenii
- 119. Clypeal hairs of A, acaci
- 120. Same as 118
- 121. Same as 118
- 122. Same as 115
- 123. Abdominal segment IV of A. baezai
- 124. Abdominal segment IV of A. peditaeniatus
- 125. Clypeal hairs of A. samarensis
- 126. Clypeal hairs of A. baezai
- 127. Anal segment of A. pseudobarbirostris
- 128. Antenna of A. pseudobarbirostris
- 129. Anal segment of A, manalangi
- 130. Antenna of A. manalangi
- 131. Prothoracic hairs 1-3 of A. peditaeniatus
- 132. Prothoracic hairs 1-3 of A. manalangi 133. Hairs on mesothorax of A. peditaeniatus
- 134. Hairs on mesothorax of A. lesteri
- 135. Clypeal hairs of A, pseudosinensis

- 136. Clypeal hairs of A. lesteri
- 137. Clypeal hairs of A. manalangi 138. Clypeal hairs of A. vanus
- 139. Abdominal segment II of A. vanus
- 140. Abdominal segment II of A. franciscoi
- 141. Abdominal segments 1-VII of A. filipinae
- 142. Abdominal segments I-VII of A. annularis
- 143. Abdominal segment II of A. filipinae
- 144. Abdominal segment 11 of A. m. flavirostris
- 145. Abdominal segment VII of A. m. flavirostris
- 146. Abdominal segment VII of A. mangyanus 147. Clypeal hairs of A. b. baisasi
- 148. Clypeal hairs of A. I. ludlowae
- 149. Clypeal hairs of A. annularis
- 150. A Same as 147
 - B Clypeal hairs of A. karwari
- 151. Prothoracic hairs 1-3 of A. annularis
- 152. Prothoracic hairs I-3 of A. philippinensis
- 153, Abdominal segment II of A, tessellatus
- 154. Abdominal segment II of A. kochi
- 155. Abdominal segment III of A. tessellatus
- 156. Abdominal segment III of A. b. baisasi
- 157. Abdominal palmate on II of A, b. balabacensis
- 158. Abdominal palmate on II of A. b. baisasi
- 159. Abdominal segment I of A. b. baisasi
- 160. Abdominal segment 1 of A. r. riparis
- 161. Prothoracic hairs 1-3 of A. kochi
- 162. Abdominal segment III of A. kochi
- 163. Prothoracic hairs 1-3 of A. maculatus
- 164. Abdominal segment III of A. maculatus
- 165. Abdominal segments III-VI of A. kolambuganensis
- 166. Abdominal segments III-VI of A. maculatus
- 167. Clypeal hairs of A. cristatus
- 168. Same as 150B
- 169. Palmate hair leaflets on III of A. karwari
- 170. Palmate hair leaflets on III of A. maculatus
- 171. Abdominal segments IV-V of A. I. ludlowae
- 172. Abdominal segment IV-V of A, v. limosus
- 173. Prothoracic hairs 1-3 of A. litoralis
- 174. Comb teeth of A. litoralis
- 175. Prothoracic hairs 1-3 of A. v. limosus
- 176. Comb teeth of A. v. limosus
- 177. A Pro- and mesothoracic pleural hairs of A. parangensis
 - Pro- and mesothoracic pleural hairs of A. indefinitus
- 178. Clypeal hairs of A. v. limosus
- 179. Clypeal hairs of A. indefinitus
- 180. Clypeal hairs of A. v. vagus
- 181. Same as 178
- 182. Prothoracic hairs 1-3 of A. indefinitus
- 183. Prothoracic hairs 1-3 of A. subpictus

INTRODUCTION

The first illustrated keys to the Anopheles mosquitoes of the Philippines were prepared by Russell and Baisas (1934, 1936). Periodically, since then keys have been produced (Simmons and Aitken, 1942; Bohart, 1945; Mendoza, 1954 a, b; Baisas and Bañez, 1957; and Baisas and Dowell. 1965).

The keys presented herein are an effort to produce simplified, well-illustrated guides to the identification of the Philippine anophelines, and have been tested extensively for the past two years on participant classes at the Malaria Eradication Training Center. The latest supra-specific concepts, especially those introduced by Reid (1968) are incorporated. Likewise, additional specific and sub-specific taxa are included.

Both the adult female and fourth instar larval stages have been dealt with in the keys. Captions for the key illustrations may be known by consulting the Table of Key Figures, page 1. The essential morphology of these forms, discussed in the succeeding sections, will enable the user to follow the keys successfully.

CLASSIFICATION OF PHILIPPINE ANOPHELES

The systematic index to the Philippine Anopheles species below follows the arrangement made by Reid and Knight (1961) and Reid (1968). It updates the checklists of Delfinado (1965) and Baisas and Dowell (loc. cit.). Unlike previous lists, here the former "groups" under subgenus Cellia are elevated to "series" which match the long-used series of the subgenus Anopheles. Furthermore, closely related species are arranged into "species groups", not "complexes" or "groups", as formerly done. With these changes the classification becomes much less complicated. Where trinomials are used in the paper, the third name indicates subspecies, i.e., Anopheles gigas formosus Ludlow. No scientific name lower than subspecies is recognized as valid.

One series name change is adopted here, i.e., Pyretophorus for Pseudomyzomyia, as suggested by Reid (1968).

The listing of the species groups to which the Philippine species belong, where applicable, serves to demonstrate the relationship of the local taxa to the Oriental anopheline fauna as a whole.

It was the opinion of Reid (1966, 1968) that A. subpictus Grassi may be absent from the Philippine Islands. Investigation of reared associated "indefinitus"

from the saline, fish ponds near Manila has shown that some of these specimens are identical to the description of "subpictus" given by Reid. Likewise, A. vagus yagus Donitz has never been reported from the Philippines since King (1932) described the insular form, A. v. limosus. It has recently been collected from Mindanao Island by the authors. Therefore, these two are included in the keys and considered to be members of the Philippine fauna.

A new subspecies of *ludlowae* has recently been described by Darsie and Ramos (1969), and named *ludlowae cabrerai*. Reference was made to it by Baisas and Dowell (loc. cit.) and Reid (1968) as a variety of *ludlowae* which bears three dark spots on the anal vein instead of the usual two. It has been included in the following keys.

SYSTEMATIC INDEX ANOPHELES OF THE PHILIPPINES

Genus Anopheles Meigen Subgenus Anopheles Meigen Anopheles series Edwards aitkenii species group acaci Baisas aitkenii James bengalensis Puri fragilis (Theobald) insulaeflorum (Swellengrebel and Swellengrebel de Graaf) lindesavi species group gigas formosus Ludlow lindesayi benguetensis King Myzorhynchus series Edwards albotaeniatus species group balerensis Mendoza eiercitoi Mendoza bancroftii species group pseudobarbirostris Ludlow barbirostris species group franciscoi Reid manalangi Mendoza vanus Walker hyrcanus species group lesteri Baisas and Hu peditaeniatus (Leicester). pseudosinensis Baisas umbrosus species group baezai Gater

Myzomvia series Christophers minimus species group filipinae Manalang mangyanus (Banks) minimus flavirostris (Ludlow) Neocellia series Christophers karwari (James) maculatus Theobald annularis species group annularis Van der Wulp philippinensis Ludlow Neomyzomvia series Christophers kochi Donitz kolambuganensis Baisas tessellatus Theobald leucosphyrus species group balahacensis baisasi Colless balabacensis balabacensis Baisas cristatus King and Baisas riparis riparis King and Baisas Pyretophorus series Edwards indefinitus (Ludlow) subpictus Grassi vagus limosus King vagus vagus Donitz ludlowae species group litoralis King ludlowae ludlowae (Theobald) ludlowae cabrerai Darsie and Ramos parangensis (Ludlow)

samarensis Rozeboom

Subgenus Cellia Theobald

The Adult Female

MORPHOLOGY OF ADULT FEMALE

It is assumed that the user is already able to distinguish not only members of the Family Culicidae from other dipterous insects, but can also differentiate anopheline from culicine mosquitoes. If not, the reader is referred to Borror and Delong (1963) for the former and to Russell et al. (1963) or Delfinado (1966) for the latter.

The description below is by no means complete, but contains sufficient background to comprehend the key points. Certain structures need to be defined in order to understand the morphology.

Scierites — The integument of insects is made up
of hardened plates called scierites, separated either
by lines, known as sutures, or membranes of various sizes. The body of the adult mosquito is composed primarily of scierites, whereas the larval
body is largely membranous.

2. Hairs and Scales — It is necessary to be able to differentiate between scales and hairs in adult mosquitoes. A hair is round, tapers from base toward apex, and it is movable, being connected to the body by a socket, called trichopore. The scales are flattened, immobile, without a trichopore, and usually widening, although the wing fringe scales are pointed, apically.

The females of Anopheles may be distinguished from those of other Philippine culicids by their long palpi, almost as long as the proboscis and by scutellum which is evenly rounded and beset by an unbroken, evenly spaced row of hairs posteriorly, see Fig. 1. Another characteristic is their living posture while at rest or taking a blood meal. Typically the body is held at a distinct angle from the resting surface and their thorax and abdomen form a straight line with the proboscis. The Head

The spherical head, seen in Fig. 2, is about as wide as long, with compound eyes occupying a large portion of each side. Projecting forward are five appendages, the median slender proboscis, above which are the two palpi, and extending from the head between the eyes, the two antennae.

The proboscis is an elongate structure about onefourth the total length of the body. The visible portion is the labium, a sheath covering the piercing stylets. It is clothed with scales, usually uniformly dark with a light colored tip, the labella. In some species the apical half is wholly or partially beset with pale or flavescent scales.

The two, five-segmented palpi are also covered with scales, which are sometimes large, giving the structures a shaggy appearance. The scalation is either entirely dark or more often with pale banding. The terminal pale band is called apical; the next one, subapical, and the intervening dark band separating them is also called the subapical dark band. The number of pale bands may vary within one species, for the subapical dark band may be missing, or in others it may be found added where it normally is absent.

The antennae are composed of two basal elements, the small scape and bulbous torus. The remaining 14 rod-like segments, or flagellomeres, are collectively called the flagellum. Each flagellar part is adorned with a whorl of hairs, excepting the last, these hairs being much longer and more numerous in the male antennae. The Thorax

The thorax (Fig. 3) consists of three segments, called prothorax, mesothorax and metathorax. In mosquitoes, the mesothorax is greatly expanded at the expense of the other two. The dorsal aspect of the thorax is made up of

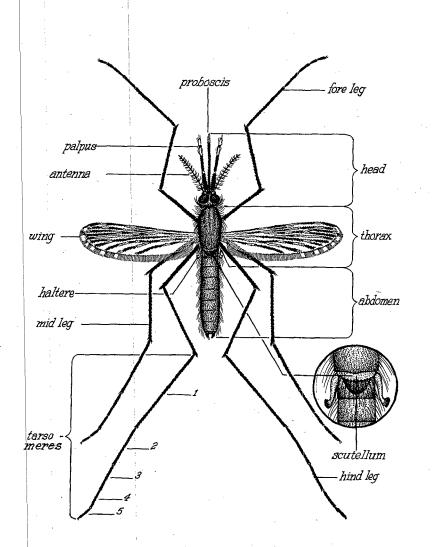


Figure 1. Portrait of *Anopheles minimus flavirostris, s*howing general morphology of a female anopheline (17x).

(Redrawn from Russell and Baisas, 1936)

the mesonotum (mn) and two smaller posterior sclerites, the scutellum (sc) and the postnotum (pn). The lateral portions of the anterior third of the mesonotum are somewhat depressed and are known as the fossae (fo). In most anophelines, scales are absent but hairs present on the dorsum of the thorax.

The lateral aspect of the thorax is known as the pleuron. It is composed of a number of sclerites which contain groups of hairs, or bristles. For the names and locations of the principal ones, see Fig. 3. The important sclerite and bristle group for the recognition of Philippine Anopheles females is the propleuron (prp) and its bristles (1).

Mosquitoes have one pair of functional wings attached to the mesothorax and one vestigial pair, called the halteres, on the metathorax. The former consists of a membrane supported by a network of veins. The veins have been named by the Comstock-Needham system in this key; see Fig. 4A for an illustration of the nomenclature. The wings are very important in the identification of Philippine anophelines. The ornamentation, especially the costal wing spots, is a salient feature. For

an understanding of the pattern and terminology used here, see Fig. 4B.

Each of the six legs is composed of nine parts, as shown in Fig. 5. Leg characters are commonly used in the identification of Philippine anophelines. Various spotting and banding of pale and dark scales will be encountered. Spots may be confused with bands, so it is necessary to make sure the pale scales extend completely around the segment.

The Abdomen

The abdomen is composed of eight visible segments designated by Roman numerals 1-VIII (Fig. 6), each consisting of a dorsal sclerite, the tergite (te), and a ventral one, the sternite (st). In most anopheline females these abdominal sclerites have few or no scales; however, in some, scales or scale tufts may be present. These dorsal and ventral sclerites are separated by a membranous integument, the pleural membrane (pm). No pleural sclerites are present on the abdomen. The terminal segments constitute the female genitalia, and the lobe-like cerci (ce) are protruding at the posterior end.

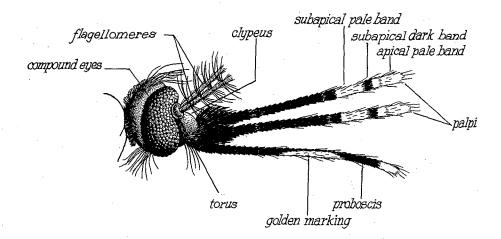


Figure 2. Head of anopheline, showing details of morphology (60x).

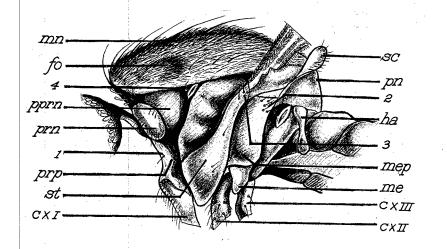


Figure 3. Thorax of anopheline showing sclerites and bristle (hair) groups (60x). Legend:

cxl coxa of prothorax cxII cxIII

coxa of mesothorax coxa of metathorax fo fossa of mesonotum ha haltere

me meron mep mesepimeron mesonotum

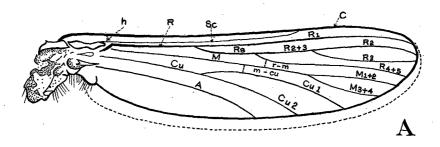
рn postnotum pronotum prn pprn postpronotum propleuron

scutellum

sternopleuron

1 propleural bristle 2 upper mesepimeral bristles

3 prealar bristles. 4 spiracular bristle



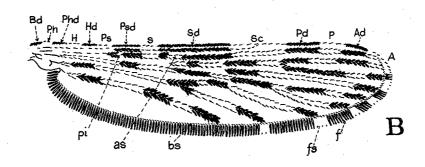


Figure 4. Wing of anopheline (44x).

	re it timeg or antepriorinte	, ,	•		
A. 1	Venation by Comstock-Nee	dham	System:		
A C Cu h	anal vein costal vein cubital vein humeral cross-vein	M m-cu R	medial vein medio-cubital cross-vein radial vein	Rs r-m Sc	radial sector vein radio-medial cross-vein subcostal vein
B. Markings on hypothetical wing:					
A Ad as Bd bs f	apical pale spot apical dark spot accessory sector pale spot basal dark spot border scales fringe scales	fs H Hd P Pd Ph Phd	fringe spot humeral pale spot humeral dark spot preapical pale spot preapical dark spot prehumeral pale spot prehumeral dark spot	pi Ps Psd S Sc Sd	pale interruption presector pale spot presector dark spot sector pale spot subcostal pale spot sector dark spot

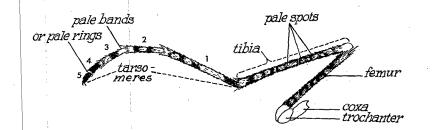


Figure 5. Leg of anopheline showing individual segments and markings (25x).

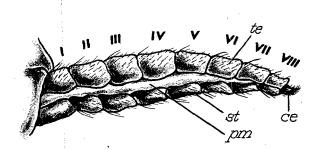
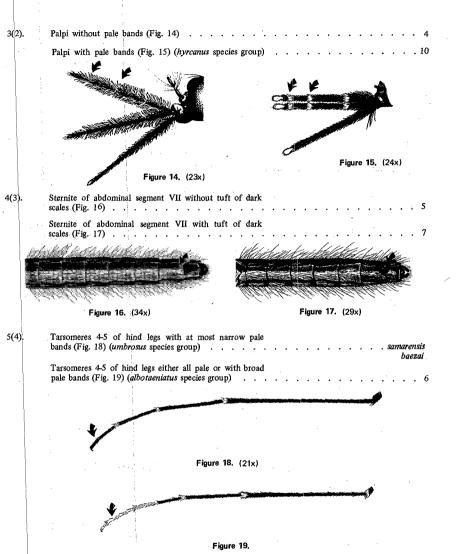


Figure 6. Abdomen of anopheline showing segments; ce - cercus; pm - pleural membrane; st - sternite; te - tergite (31x).

Illustrated Key to Adult Females of Philippine Anopheles

1.	Costal vein of wing with three or fewer pale spots, or if more than three, then sector pale spot absent (Fig. 7) (Subgenus Anopheles)	2
	Costal vein of wing with four or more pale spots, sector pale spot always present (Fig. 8) (Subgenus Cellia)	
	and the state of t	Figure 7. (33x)
	A COLUMN TO THE PARTY OF THE PA	Figure 8. (42x)
2(1).	Femora of forelegs distinctly swollen in basal half (in dried specimens swollen portion usually collapsed and with longitudinal depression) (Fig. 9); palpi shaggy in appearance sometimes only in basal half (Fig. 10) (Myzorhynchus series)	3
	Femora of forelegs not swollen (Fig. 11), or only slightly (Fig. 12), in basal half; palpi not shaggy (Fig. 13) (Anopheles series)	
	,	11/1/11
-	Figure 9. (66x)	A CONTRACTOR OF THE PARTY OF TH
		musuam ·
Ì	ALTO SHOW	Figure 10. (23x)
	Figure 11. (34x)	
Figure	12. (51x)	Figure 13. (17x)



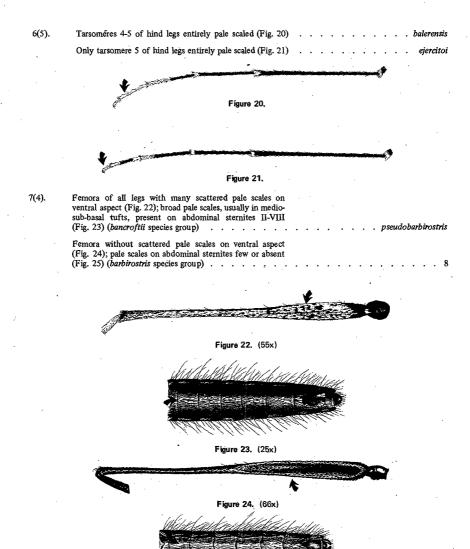


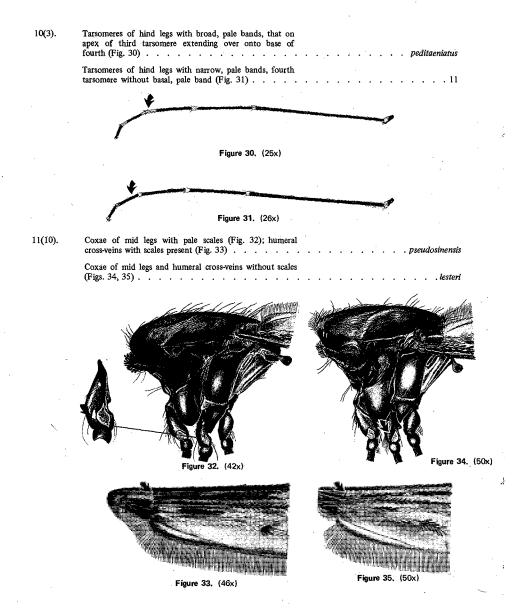
Figure 25. (21x)

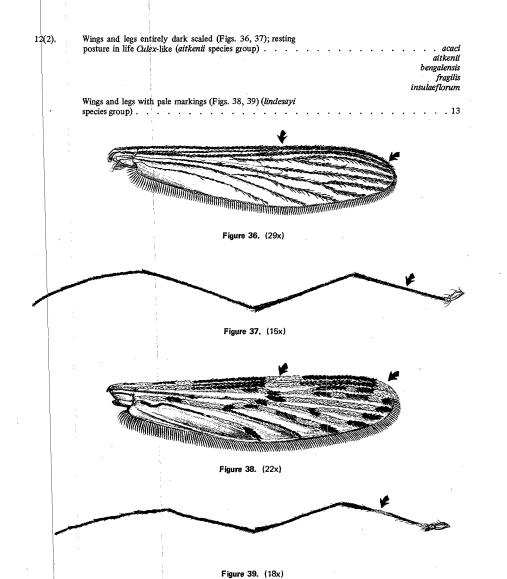
Figure 27. (26x)



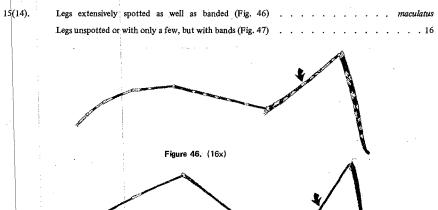
Figure 28, (21x)

Figure 29. (21x)



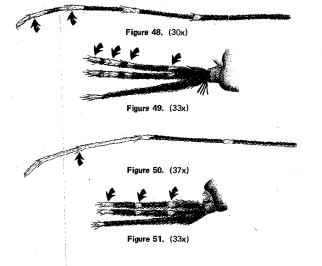


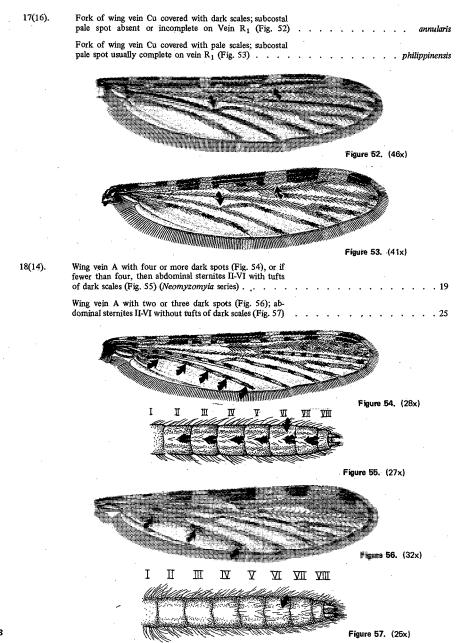
	·
13(12).	Femora of hind legs with broad, pale band at about the middle (Fig. 40); only apical pale spot present on costal vein of wing (Fig. 41)
	Femora of hind legs without pale band at about the middle (Fig. 42); costal vein with four or five broad, pale spots (Fig. 43)
	· · · · · · · · · · · · · · · · · · ·
	Figure 40. (29x)
	1 iguit 40. (23x)
	Figure 41. (25x)
	A
	Figure 42. (25x)
	Figure 43. (22x)
14(1).	Last tarsomere on hind legs entirely pale (Fig. 44) (Neocellia series)
	Last tarsomere of hind legs either entirely, or at least partially, dark scaled (Fig. 45)
•	
	Firm 44 (24)
	Figure 44. (21x)
\	Figure 45. (23x)

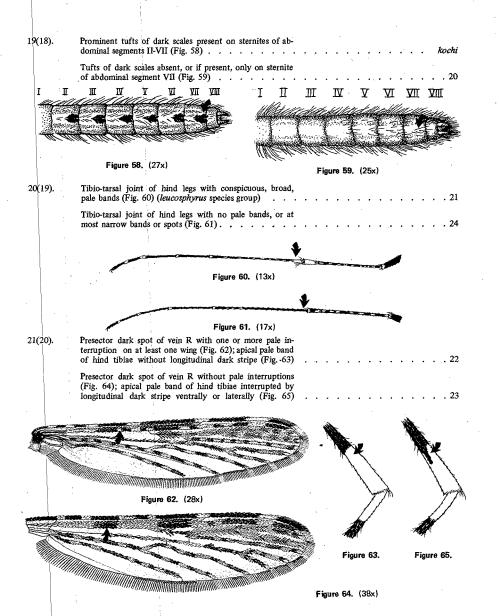


16(15). Hind tarsomeres 3-5 with broad, pale bands, only tarsomere 5 all pale (Fig. 48); palpi with four pale bands (Fig. 49).

Figure 47. (23x)







Basal-most part of presector dark spot of vein R usually extending no farther than level of presector pale spot, but sometimes to center of humeral dark spot of costa (Fig. 66); tarsomere 4 of hind legs with prominent basal pale band,

Basal-most part of presector dark spot of vein R extending well into level of humeral dark spot of costa, sometimes even to humeral pale spot (Fig. 68); tarsomere 4 of hind legs without basal pale band or with very narrow one (Fig. 69)

..



Figure 66. (28x)



Figure 67. (13x)



Figure 68, (38x)



Figure 69. (22x)

Costal vein nearly always with prehumeral pale spot (Fig. 71); proboscis/fore-femur ratio greater than 0.90.

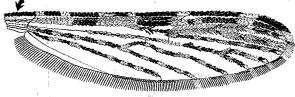


Figure 70. (38x)



Figure 71. (38x)

24(20). Tergites of abdominal segments with very few or no scattered, broad, pale scales (Fig. 72); apical half of proboscis golden, excepting narrow preapical, dark band

> Tergites of abdominal segments with many scattered, broad, pale scales (Fig. 74); proboscis usually with some golden scales, but confined to narrow preapical band or patch (Fig. 75)



Figure 72. (34x)



kolambuganensis

Figure 74. (34x)



Figure 73. (27x)



Figure 75. (33x)

25(18)	Fork of wing vein Cu with dark scales (Fig. 76) (Myzomyia series; minimus species group)
	Fork of wing vein Cu with pale scales (Fig. 77) (Pyreto-phorus series)
	Figure 76. (42x)
	Figure 77. (38x)
26(25).	Wing vein A with three dark-scaled areas, fringe opposite this vein pale (Fig. 78); subapical dark band of palpi subequal in length to subapical pale band (Fig. 79) , filipinae
	Wing vein A with only two dark areas, distal one long, basal one short, fringe opposite this vein dark (Fig. 80); subapical dark band of palpi shorter than subapical pale band (Fig. 81)
	Figure 78. (40x) Figure 79. (33x)
•	Figure 81. (32x)

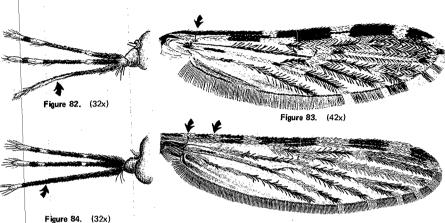
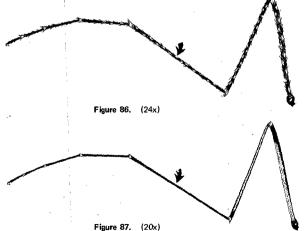
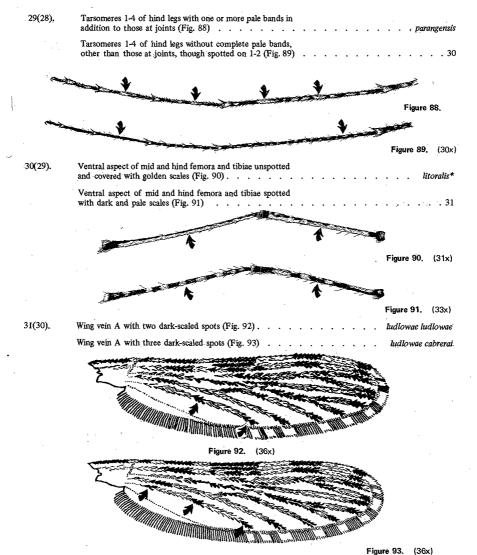
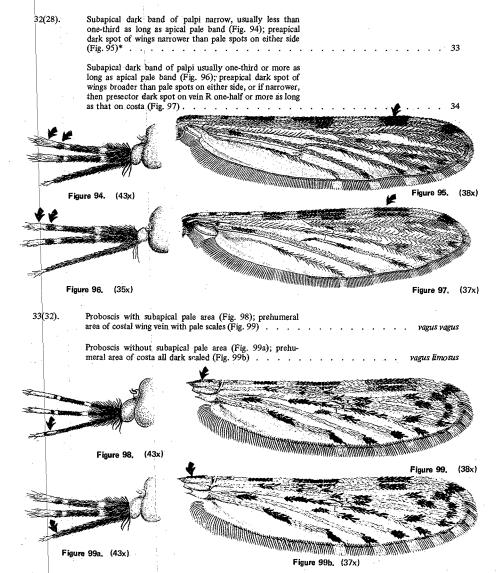


Figure 85. (38x)





*Since A. sundaicus Rodenwaldt may occur in the Philippines, it would key to literalis' in this table. It may be distinguished from literalis by the absence of the prehumeral pale spot on the wing.



*Second character does not apply to most of vagus vagus examined.

34(32).	Subapical pale band of palpi one-third or less as long as subapical dark band (Fig. 99c)
	Subapical pale band of palpi usually one-half or more as long as subapical dark band (Fig. 99d) indefinitus



Figure 99c. (30x)



Figure 99d. (35x)

The Fourth Instar Larva

MORPHOLOGY OF THE LARVA

The anopheline larvae are readily distinguished from other kinds of mosquito larvae by the absence of an elongate siphon on the eighth abdominal segment, and the presence of palmate hairs (Hair I) on some or all of abdominal segments I-VII. The attitude of the live anopheline larvae when at rest on the surface of the water is characteristic, for they lie parallel to the top, whereas, most culicine larvae lie at a distinct angle to the surface.

The body of the mosquito larva is largely membranous, and beset by sclerites, such as the head capsule, abdominal tergal plates, spiracular apparatus and anal saddle. There are also numerous hairs attached both to the sclerites and to the membranes. The study of hairs (setae) is called chaetotaxy and the complete chaetotaxy of Anopheles litoralis King is depicted in Figs. 100-103. The nomenclature of Belkin (1951) is employed for the larval hairs. The morphology of individual hairs, shown in Fig. 104, is singularly important in identification of larvae.

The larva is divided into three distinct body regions; the head, somewhat flattened; the thorax, consisting of three fused segments, each indicated by its own set of hairs; and the abdomen, with eight obvious segments and two markedly modified structurally.

There are four larval instars, each separated by a moulting of the skin (ecdysis). A rapid, morphological identification of the four instars may be made by using the following simple key:

a.	Egg breaker present on frontoclypeus First Instar
a.	Egg breaker absent b
	b. Hair 16 of maxillary palpus
	absent Second Instar
ł	bb. Hair 16 of maxillary palpus presentc
	c. Imaginal eye absent Third Instar
	cc. Imaginal eye present Fourth Instar

The fourth instar is obviously the largest in size, but unless one is familiar with relative sizes of the species, it may be a poor character to follow. For instance, the third instar of Anopheles manalangi Mendoza may be larger than the fourth instar larva of Anopheles filipinae Manalang. The accompanying key is based on the fourth instar, but in most instances may also be used to identify the third instar.

The mature fourth instar larvae (pre-pupae) can be recognized by the appearance of the pupal hairs under their skin.

The following detailed description applies to the fourth instar anopheline larva.

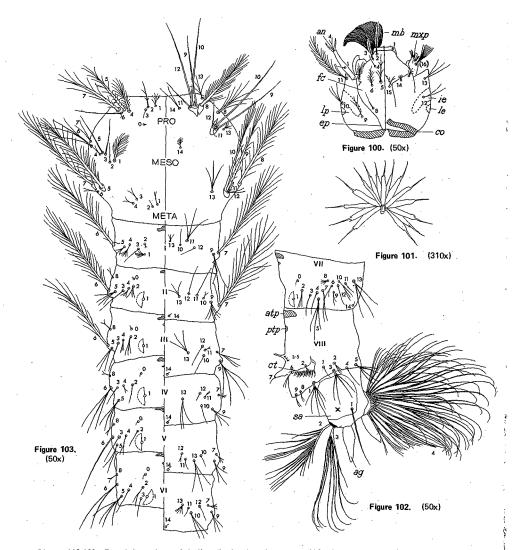
Head

The head is composed of a completely sclerotized capsule, Fig. 100. Three major plates are visible on the head, the frontoclypeus (fc), dorsally, and the two lateral plates (lp) which meet on the ventral side. A prominent line known as the epicranial suture (ep) separates the frontoclypeus from the lateral plates. Anteriorly are the antennae (an) and mouthparts. The antenna bears a hair on the shaft and several at its terminus. In some species the shaft hair (No. 1) is large, multibranched and located near the middle of the shaft. while in others it is small, simple and positioned in the basal one-third. The most conspicuous structures of the mouthparts are the mouth brushes (mb) terminal in position. The maxillary palpi (mxp) are prominent lobes just medio-ventral to the antennae, which bear subapical hairs (No. 16 of some authors: see Christophers, 1933 and Baisas and Dowell, 1965), On each lateral plate are found two eyes, a prominent imaginal eve (ie) and small, posterior, larval eve (le). The posterior border of the head is heavily sclerotized and called the collar (co). In early instars this collar is much

Some of the 15 pairs of head hairs are of major importance in identification. The inner and outer clypeal hairs, head hairs 2 and 3, are widely used. The clypeal hairs may be simple, forked at tip, frayed, variously branched or dendritic, a term applied to branching which appears like the branches of a tree; see Fig. 104, A-D, L, M. The outer clypeal hairs are often difficult to see because they overlie the mouth brushes. It helps to view them with the 43x objective of a compound microscope. The same power will be necessary to see the frayings, or minute side branches. of the clypeal hairs. Less often used are the posterior clypeal and the sutural and transsutural hairs hairs 4, 8 and 9, respectively. Hair 16 on the maxillary palpi is not named by Belkin, but it is included for convenience since it is useful in larval instar recognition.

Thorax

The thorax is the thickest part of the body, and composed of three segments, the prothorax, mesothorax and metathorax. They are completely fused into one large body region, but the evidence of segmentation is found in the chaetotaxy, for there are three distinct set of hairs; see Fig. 103.



Figures 100-103. Fourth instar larva of *A. litoralis*, showing chaetotaxy. 100. Head, dorsal — left, ventral — right; 101. Palmate hair of abdominal segment IV; 102. Terminal segments of abdomen; 103. Thorax and abdominal segments I-VI, dorsal — left, ventral — right. *Legend*:

- anal gill ep - epicranial suture mxp - maxillary palpus fc - frontoclypeus - antenna ptp - posterior tergal plate ie - imaginal eye - anterior tergal plate - anal saddle - collar le - larval eye co PRO - prothorax - comb teeth lp - lateral plate MESO - mesothorax mb - mouth brush META - metathorax

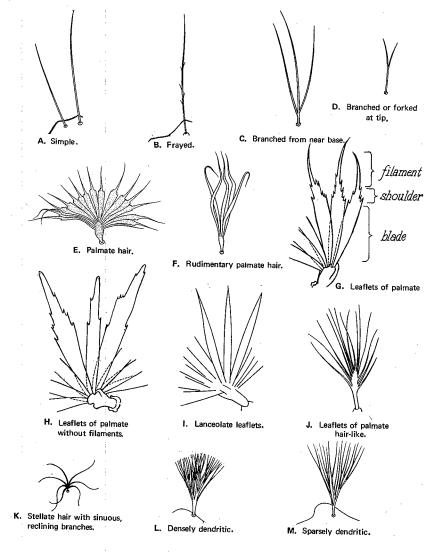


Figure 104. Morphology of Hairs.

Certain thoracic hairs are used as diagnostic characters. The submedian prothoracic hairs 1, 2 and 3 show differences not only in the number of branches, but also the configuration of their bases. Sometimes their trichopores are set into sclerotized tubercles, which at times are joined together. Mesothoracic hair 4 (hair 5 of Puri, 1960, and Baisas and Dowell, loc. cit.) in one species has reclining, sinuous branches which form a star shape (stellate), whereas normally the branches are erect (Fig. 104. C, K). The pleural hair groups, hairs 9-12, on the ventral side of each thoracic segment are important because they differ in the number of branches among the various species. They are distinctive because each group of four hairs arises from a common tubercle. Metathoracic hair 3 may have hair-like or leaf-like branches. In the latter case it is known as the metathoracic palmate hair.

Abdomen

Following the interpretation of Snodgrass (1959) and Puri (loc. cit.), the abdomen consists of 10 segments of which the first seven are similar in composition; see Fig. 102. The eighth is modified to bear the spiracular apparatus, and the latter is composed of elements of both the eighth and ninth abdominal segments. Two sets of hairs, as interpreted by Belkin, on the eighth segment support the opinion that actually two segments are represented, VIII and 1X. The terminal segment is the tenth, which ends in the anus, carries large dorsal and ventral tufts of hairs and possesses four anal gills (ag), thin-walled, tracheated lobes or papillae.

Each of the first eight abdominal segments has a sclerotized plate dorsally, called the anterior tergal plate (atp). Posterior to it on some segments are one or more smaller sclerites known as posterior tergal plates (ptp). The tergal plates may be difficult to see if the alimentary canal is filled with dense material. The tenth segment also has a large dorsal plate, called the saddle (sa). The larvae belonging to Myzomyia series have large anterior tergal plates, sometimes almost covering the entire dorsal surface of the segment.

The normal complement of hairs on the abdominal segments is 15 hairs, Nos. 0-14; however, VIII, IX and X have fewer; see Figs. 103, 102.

Certain abdominal hairs have decided importance as recognition characters. The palmate hairs (Hairs No. 1) are salient features on I-VII. Figure 104, E-J, show the various forms. The palmates may have leaf-like, flattened branches (E, G, I) or hair-like parts (J). The usual leaflet has a basal flat part, the blade, terminal, attenuated portion, the filament, and a section in between where a series of notches occur, usually with a more or less flattened area, known as the shoulder (G). Some species have the notches but no distinct shoulders. Others are without the attenuated filament, the end being blunt (H). A well developed palmate will usually have 10 or more leaflets like those just described (E). A rudimentary palmate hair (F) commonly has two to eight leaflets, lanceolate in shape, i.e., not very wide and with few or no notches.

Next in importance are the lateral hairs (Hairs No. 6) on abdominal segments III-VI; see Fig. 103. These are usually the longest hairs on the segment. Their length in relation to each other, number and mode of branching are useful characters. The antepalmate hairs (Hairs No. 2) particularly on abdominal segment VII are utilized for identification.

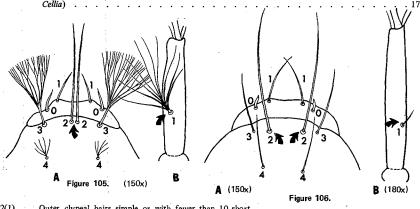
The spiracular apparatus on VIII (Fig. 127) consists of a central depression, the respiratory fossa, in which the spiracles are located. The spiracles constitute the only point of intake for atmospheric air required by the larva in respiration. Surrounding the fossa are four protective structures, the fan-shaped plate (ap) anteriorly, the two lateral papillae (lpa), and the posterior, concave scoop (sc). The anterior plate sometimes bears a long, thin, appendage, the stigmal club (scl), which extends posteriorly between the spiracles and overlies the scoop. The scoop has two lateral plates, forming the side walls, and a median plate, heavily sclerotized anteriorly.

On either side of the spiracular apparatus are strongly sclerotized plates, the combs, or pecten. In the Philippine anopheline larvae the posterior edge of the comb carries a number of teeth, usually several long and many more distinctly short. However, they may be subequal or have graduated lengths. Their bases ordinarily have fine serrations.

Illustrated Key to Fourth Instar Larvae of Philippine Anopheles*

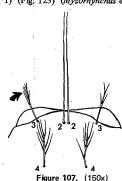
1. Inner clypeal hairs (Hair 2) close together, the distance between their bases less than the distance between the bases of the inner and outer (Hair 3) clypeal hairs (Fig. 105A); antennal hair (Hair 1) multibranched, often large (Fig. 105B) (Subgenus Anopheles)

Inner clypeal hairs widely separated, the distance between their bases usually more than the distance between the bases of the inner and outer clypeal hairs (Fig. 106A); antennal hair simple and minute (Fig. 106B) (Subgenus Cellia)

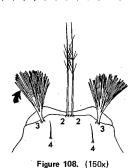


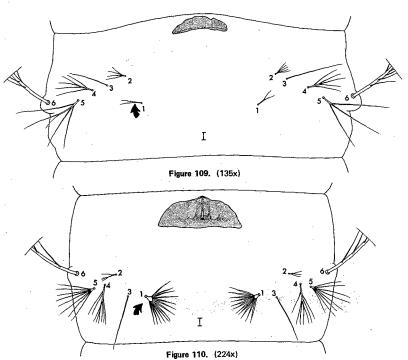
2(1). Outer clypeal hairs simple or with fewer than 10 short, lateral branches (Fig. 107) (Anopheles series).

Outer clypeal hairs dendritic, with 20-60 branches (Fig. 108), or if fewer than 20, then abdomen without palmate hairs (Hair 1) (Fig. 123) (Myzorhynchus series).



*Larvae of A. balerensis and A. ejercitoi unknown.







(Fig. I12)

Figure 111. (350x)



lindesayi benguetensis

Figure 112. (350x)

5(3). Inner clypeal hairs simple, their bases closer together than distance to outer clypeals (Fig. 113); lateral hairs of abdominal segment III with three to nine branches (Fig. 114) . insulaeflorum Inner clypeal hairs branched or frayed, and their bases not much closer together than the distance to the outer clypeals (Fig. 115); lateral hairs of abdominal segment III with 20-50 branches (Fig. 116) Figure 113. (90x) (210x) Figure 114, (120x) Figure 116. (110x) (89x) Figure 115. (250x) 6(5). Middle area of inner clypeal hairs with fine side frayings, stem single or two- to three-branched (Fig. 117) Middle area of inner clypeal hairs without such fine frayings, stem with two to 14 branches (Fig. 118)

Figure 118.

(220x).

Figure 117.

(230x)

33

7(6).

Inner clypeal hairs with branches well separated and easily distinguishable, not bent down, with fewer than nine branches (Fig. 120)

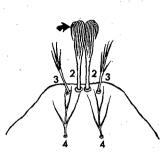
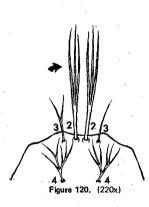


Figure 119. (220x)



8(7). Inner clypeal hairs with two or three branches, all beginning about one-fourth of the distance from base (Fig. 121) aitkeni

Inner clypeal hairs usually with four to seven branches, if two- or three-branched, then branches beginning one-third or farther from base (Fig. 122)

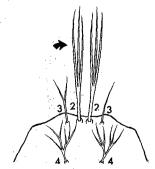


Figure 121. (220x)

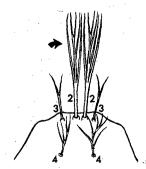
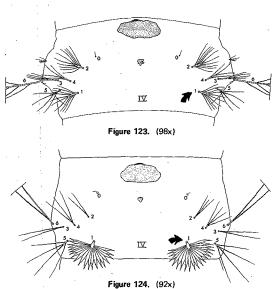
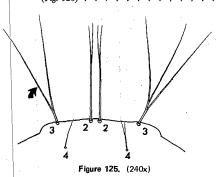


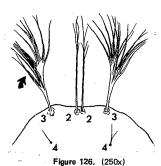
Figure 122. (250x)

2).	Palmate hairs undeveloped on thorax and abdomen,
	branches hair-like (Fig. 123) (umbrosus species group)
	Palmate hairs well developed on at least some abdominal
	segments, branches broad, leaf-like (Fig. 124)





10(9).



11(9).	antennal shaft swollen basally, narrowing only beyond hair I (Fig. 128) (bancroftii species group)		pseudobarbirostris
	Stigmal club absent (Fig. 129); antennal shaft gradually narrowing apically (Fig. 130)		12
so	Figure 127. Dorsal-left; lateral-right (89x). ap anterior plate lpa lateral papilla sc scoop scl stigmal club		
Figure 12	29. Dorsal view (102x)	Figure 128. F (190x)	igure 130. (165x)
12(11).	Prothoracic hair 1 simple or with two or three short branches at tip (Fig. 131) (hyrcanus species group)		13
	Prothoracic hair 1 with four to 15 branches (Fig. 132) (barbirostris species group)		15



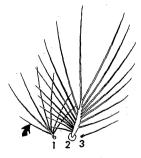
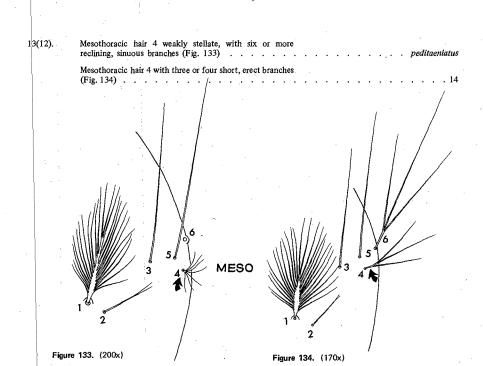
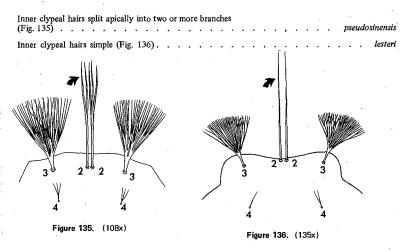
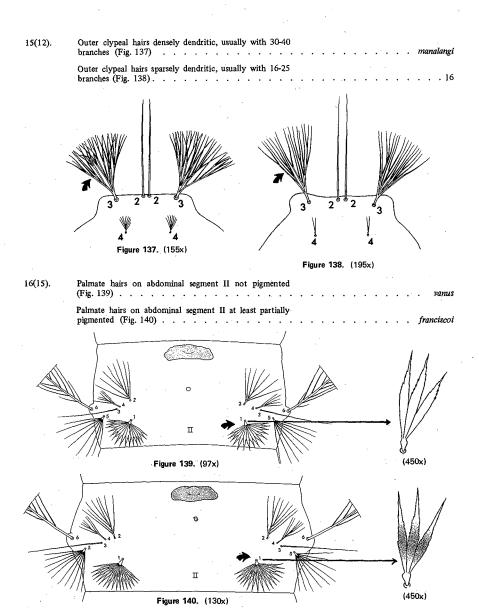


Figure 132. (220x)



14(13).





Anterior tergal plates on abdominal segments large (Fig. 141) (Myzomia series; minimus species group)
Anterior tergal plates on abdominal segments small (Fig. 142)
II
Figure 141 (45v)
Figure 141. (45x) Figure 142. (39x) Anterior tereal plate on abdominal segment II not indented.
Anterior tergal plate on abdominal segment II not indented, usually convex, posteriorly (Fig. 143),
Anterior tergal plate on abdominal segment II not indented,
Anterior tergal plate on abdominal segment II not indented, usually convex, posteriorly (Fig. 143)
Anterior tergal plate on abdominal segment II not indented, usually convex, posteriorly (Fig. 143). Anterior tergal plate on abdominal segment II indented posteriorly (Fig. 144)

Figure 144. (145x)

17(1).

18(17).

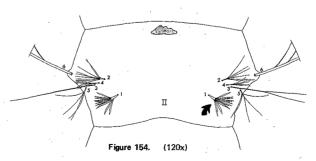
B (210x)

Figure 150.

Figure 149. (160x)

A (230x)

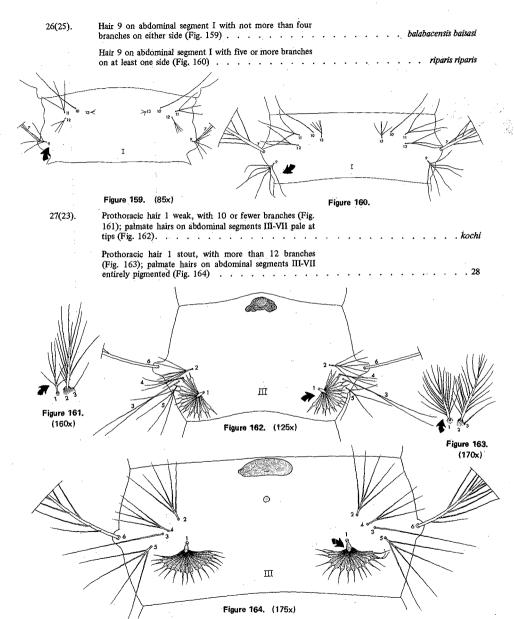
Tubercles of prothoracic hairs 1 and 2 fused and deeply pigmented (Fig. 151)	aris
Tubercles of prothoracic hairs 1 and 2 separated and pale or transparent (Fig. 152)	ısis
Figure 151. (135x) Figure 152. (175x)	
Palmate hairs of abdominal segment II not fully developed, hair-like or at most with thin leaflets (Fig. 153)	24
Palmate hairs of abdominal segment II fully developed with clearly differentiated leaflets (Fig. 154)	27
Figure 153. (126x)	

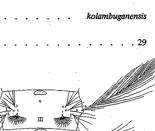


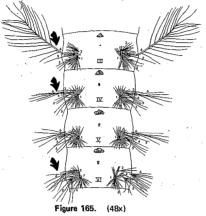
	· · · · · · · · · · · · · · · · · · ·
24(23).	Leaflets of palmate hairs on abdominal segments III-VII lanceolate; lateral hairs of abdominal segment III with fewer than 10 branches (Fig. 155)
:	Leaflets of palmate hairs on abdominal segments III-VII with distinct shoulders and filaments, lateral hairs of abdominal segment III with more than 10 branches (Fig. 156)
	4 3 6
	ш
11/1/2	Figure 155. (175x)
(160x)	Figure 156. (105x) (110x)
25(24).	Abdominal segment II with palmate hairs moderately developed, usually with thin lanceolate leaflets (Fig. 157) balabacensis balabacensis
	Abdominal segment II with palmate hairs weakly developed, branches hair-like or only slightly flattened (Fig. 158)

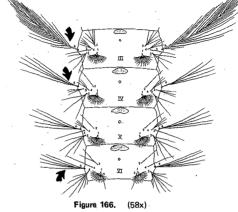
Figure 158. (570x)

Figure 157. (520x)









29(28). Posterior clypeal hairs (Hair 4) with unusually long, lateral branches, similar to outer clypeal hairs (Fig. 167) . . .

Posterior clypeal hairs long or short, but without long lateral branches, usually single (Fig. 168) .

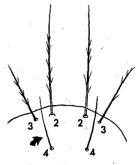


Figure 167, (180x)

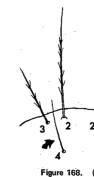


Figure 168. (210x)

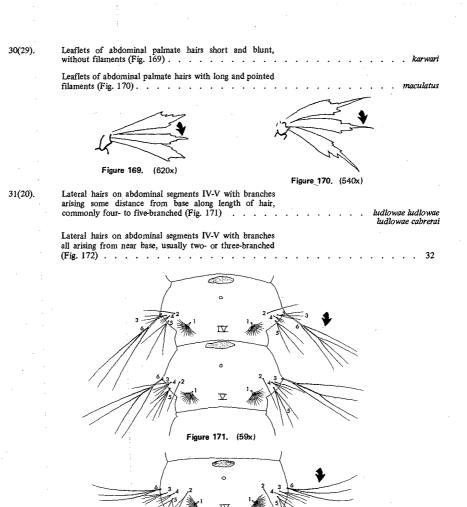
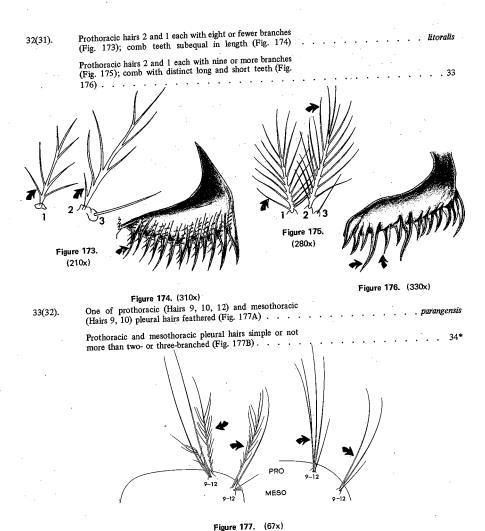
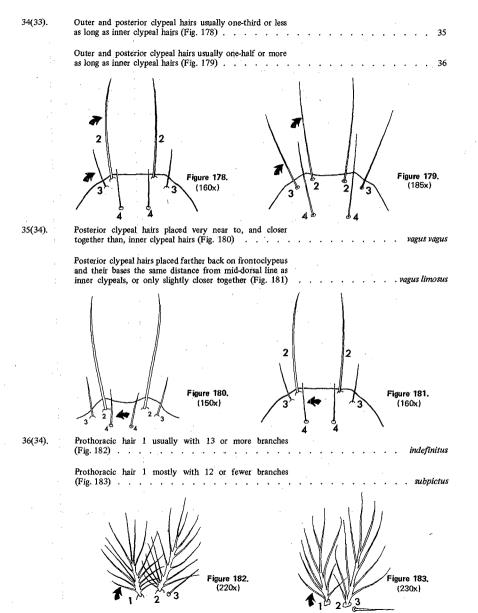


Figure 172. (64x)



^{*}Since A. sundaicus Rodenwaldt may occur in the Philippines, its larvae will come out in this key to the second part of couplet 33. It may be distinguished from vagus limosus and indefinitus by mesothoracic hair 4, which in sundaicus has three branches from near base, and in the latter two, is double, or if three-branched, the third arises from halfway along the hair.



REFERENCES:

- Baisas, F.E. and F.L. Banez. 1960. Arthropods of medical importance. Inst. Malariol., Div. Malaria, Bur. Dis. Control, Dept. Hlth., Manila, Philippines, 105 pp (mimeo.).
- ____and F.H. Dowell. 1965. Keys to the adult female and larval anopheline mosquitoes of the Philippines. Jour. Med. Ent. 4(1):11-23.
- Belkin, J.N. 1951. A revised nomenclature for the chaetotaxy of the mosquito larva. Amer. Mid. Nat. 44:678-698.
- Bohart, R.M. 1945(?) A synopsis of the Philippine mosquitoes. U.S. Naval Med. Res. Unit No. 2, Navmed 580, 88 pp.
- Borror, D.J. and D.M. Delong. 1963. An introduction to the study of insects. New York, Holt, Rinehart and Winston, 819 pp.
- Christophers, S.R. 1933. Fauna of British India, Diptera, Family Culicidae, Tribe Anophelini, Vol. IV, London, Taylor and Francis, 371 pp.
- Colless, D.H. 1956. The Anopheles leucosphyrus group. Trans. Roy. Ent. Soc. London. 108(3):37-116.
- Darsie, R.F. Jr. and A.C. Ramos. 1969. Manual of malaria entomology. 2nd Ed. Manila, Malaria Eradication Training Center. 142 pp.
- Delfinado, M.D. 1966. The culicine mosquitoes of the Philippines, Tribe Culicini (Diptera, Culicidae). Mem. Amer. Ent. Inst. No. 7, 252 pp.
- _____, G.B. Viado and L.T. Coronel. 1962. A checklist of Philippine mosquitoes with a larval key to genera (Diptera, Culicidae). Phil, Jour. Sci. 91(4):433-457.
- King, W.V. 1932. The Philippine Anopheles of the rossi-ludlowae group. Phil. Jour. Sci. 47(3):305-342.
- and F.E. Baisas. 1936. A new species and a new variety of Philippine Anopheles related to Anopheles leucosphyrus Donitz. Proc. Ent. Soc. Wash. 38(5):79-89.
- Mendoza, J.B. 1948. Two more Philippine Anopheles in the Myzorhynchus series. Monthly Bull. Bur. Health. 33(3):171-184.
- _____1954a. Pictorial key to the adults (females) of Philippine Anopheles. Publ. Hith. Res. Lab., Div. Malaria, Bur. Dis. Control, Dept. Hith., Manila, Philippines.
- ______1954b. Pictorial key to Philippine anopheline larvae. Publ. Hlth. Res. Lab., Div. Malaria, Bur. Dis. Control, Dept. Hlth., Manila, Philippines.
- Puri, I.M. 1960. Synoptic tables for the identification of the full-grown larvae of the Indian anopheline mosquitoes. Indian Hlth. Bull. 16, 7th Ed., 104 pp.
- Reid, J.A. 1966. A note on Anopheles subpictus Grassi and A. indefinitus Ludlow (Diptera: Culicidae) Jour. Med. Ent. 3(3-4):327-331.
- ————— 1968. Anopheline mosquitoes of Malaya and Borneo. Studies Inst. Med. Res., Malaysia, No. 31, 520 pp.
- and K.L. Knight. 1961. Classification within the subgenus Anopheles (Diptera: Culicidae). Ann. Trop. Med. & Parasit. 55(4):474-488.

Russell, P.F. and F.E. Baisas. 1934. A practical illustrated key to larvae of Philippine Anopheles. Phil. Jour. Sci. 55(4):305-336.

1936. A practical illustrated key to adults of Philippine Anopheles. Phil. Jour. Sci. 59(1):15-64.

L.S. West, R.D. Manwell and G. MacDonald. 1963. Practical malariology, 2nd Ed. London, Oxford Univ. Press, 750 pp.

Simmons, J.S. and T.H.G. Aitken. 1942. The anopheline mosquitoes of the northern half of the Western Hemisphere and of the Philippine Islands, U.S. Army Med. Bull. No. 59, 213 pp.

Snodgrass, R.E. 1959. The anatomical life of the mosquito. Smiths. Misc. Coll. 139(8):1-87.

